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## **PCT**

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	ed proteins or chemicals in fungal host cells which comprise modulation. The amino acid and nucleic acid sequence of hbrA is provided.
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### Description

#### **HYPHAL GROWTH IN FUNGI**

#### Field of the Invention

The present invention generally relates to hyphal growth in fungi and in particular describes the modulation of genes associated with hyphal growth in filamentous fungi. The present invention provides methods and systems for the production of proteins and/or chemicals from filamentous fungi which comprise modulation of genes associated with hyphal growth.

#### Background of the Invention

While the number of fungal species described is approximately 64,000, it is estimated that over one million species exist making this a diverse group of organisms. About 90% of fungi grow in the form of a radiating system of branching hyphae known as the mycelium. This mode of growth reflects a different life style from unitary organisms such as yeasts, with distinct advantages for advancing over surfaces and penetrating substrata (Carlile, 1994, The Growing Fungus, ed. Gow, N.A.R. & Gadd, G.M., Chapman & Hall, pp.3-19). To date very few genes have been characterized which effect fungal branching. The most characterized gene is cot1 isolated from the fungus *Neurospora crassa*. Cot-1 is a temperature sensitive mutation leading to hyperbranching and the sequence, whose function is unknown, appears to encode a cAMP dependent protein kinase (Yarden et al. 1992, EMBO J. 11:2159-2166).

Filamentous fungi find industrial importance as producers of antibiotics, enzymes, fine chemicals and food (<u>Aspergillus</u>: 50 Years On (1994) vol 29, ed S.D. Martinelli & J.R. Kinghorn pp. 561-596). There remains a need in the art for improved methods of producing proteins in filamentous fungus. Filamentous fungus are also known pathogens of plants and animals. Therefore, understanding the genetic basis of fungal growth will provide insight regarding possible anti-fungal therapies.

#### Summary of the Invention

The present invention is based, in part, upon the discovery of *Aspergillus* genes that are associated with fungal morphology and in particular with hyphal branching. A linear relationship between the degree of hyphal branching (measured as hyphal growth unit length) and culture viscosity in the fermentor (as measured by torque exerted on the rheometer impeller) has been observed. Isolation of hyper branching fungal mutants and identification of genes associated with fungal hyper branching provides a means for modulating fungal morphology thereby providing a means for controlling viscosity and improving fermentor performance.

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The present invention is also based, in part, upon the identification of an *A. nidulans* mutant for the production of HbrA (the mutant being referred to herein as HbrA2) which exhibits a hyperbranching phenotype at the restrictive temperature, 42°C. The mutation HbrA2 does not appear to affect growth of *A. nidulans* at 26°C, but results in a hyperbranching, restricted growth phenotype at 42°C. The HbrA2 mutant comprising the heterologous nucleic acid encoding the *M.meihei* protease was able to secrete the protease at 26°C. The HbrA2 mutant was unable to secrete the protease at 37°C but was able to secrete the endogenous alpha amylase at temperatures greater than 37°C. The present invention provides the amino acid, HbrA, and nucleic acid sequence for *hbrA* and methods for producing heterologous protein or chemicals in fungi by modulating the expression of proteins associated with hyphal growth, such as HbrA.

Accordingly, the present invention provides an isolated protein associated with hyphal growth in fungi having at least 70%, at least 75%, at least 80%, at least 85%, at least 90% or at least 95% identity to the amino acid sequence as disclosed in SEQ ID NO:2. In one embodiment, the protein associated with hyphal growth is HbrA which has the amino acid sequence as disclosed in SEQ ID NO:2. The present invention provides polynucleotides encoding the amino acid having the sequence as shown in SEQ ID NO:2 as well as polynucleotides having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90% or at least 95% identity to the polynucleotide having the sequence as shown in SEQ ID NO:1. In one embodiment, the polynucleotide is capable of hybridizing to the polynucleotide having the sequence as shown in SEQ ID NO:1 under conditions of intermediate to high stringency, or is complementary to the polynucleotide having the sequence as shown in SEQ ID NO:1. In another embodiment, the polynucleotide has the nucleic acid sequence as disclosed in SEQ ID NO:1. The present invention also provides host cells and expression vectors comprising a polynucleotide encoding SEQ ID NO:2

In one embodiment, the host cell is a fungus and in another is a filamentous fungus including *Aspergillus, Trichoderma, Mucor* and *Fusarium*. In yet a further embodiment, the *Aspergillus* species includes, but is not limited to, *A. niger*, *A. nidulans*, *A. oryzae* and *A. fumigatus*.

The present invention also provides a method for producing a desired protein in a fungus comprising the step of culturing a recombinant fungus comprising a polynucleotide encoding the desired protein under conditions suitable for the production of said desired protein, said recombinant fungus further comprising a polynucleotide encoding a protein associated with hyphal growth in said fungus said protein associated with hyphal growth

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having at least 70%, at least 75%, at least 80%, at least 85%, at least 90% or at least 95% identity to the amino acid sequence as disclosed in SEQ ID NO:2. In one embodiment, the polynucleotide encoding a protein associated with hyphal growth is homologous to said fungus and is present in amounts greater than found in the naturally occurring fungus. In another embodiment, the polynucleotide encoding a protein associated with hyphal growth is heterologus to said fungus and has been recombinantly introduced into said fungus. The method may further comprise the step of recovering said desired protein.

In another aspect of the present invention, it may be desirable to down regulate expression of the protein associated with hyphal growth in order to reduce culture viscosity. Accordingly, the present invention provides a method for producing a desired protein in a fungus comprising the step of culturing a recombinant fungus comprising a polynucleotide encoding the desired protein under conditions suitable for the production of said desired protein, said recombinant fungus comprising a mutation in an endogenous nucleic acid encoding a protein associated with hyphal growth said mutation resulting in the inhibition of the production by said fungus of the protein associated with hyphal growth.

In one embodiment, the polynucleotide encoding a protein associated with hyphal growth in said fungus comprises a replicating plasmid. In another embodiment, the polynucleotide encoding a protein associated with hyphal growth in said fungus is integrated into the fungal genome. In yet a further embodiment, the protein associated with hyphal growth has the amino acid sequence as shown in SEQ ID NO:2.

In yet a further embodiment of the present invention, the polynucleotide encoding a protein associated with hyphal growth has at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90% or at least 95% identity to the polynucleotide having the sequence as shown in SEQ ID NO: 1, or is capable of hybridizing to the polynucleotide having the sequence as shown in SEQ ID NO:1 under conditions of intermediate to high stringency, or is complementary to the polynucleotide having the sequence as shown in SEQ ID NO:1. In another embodiment, the polynucletoide has the nucleic acid sequence as shown in SEQ ID NO: 1.

The present invention also provides a method for producing a recombinant fungus comprising a polynucleotide encoding a protein associated with hyphal growth comprising the steps of obtaining a polynucleotide encoding said protein associated with hyphal growth; introducing said polynucleotide into said host cell; and growing said host cell under conditions suitable for the production of said protein associated with hyphal growth. In one embodiment of this method, the host cell is a fungus. In another embodiment, the filamentous fungus includes Aspergillus, Trichoderma, Mucor and Fusarium species. In yet

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another embodiment, the Aspergillus species includes A. niger, A. nidulans, A. oryzae and A. fumigatus. In one embodiment, the polynucleotide has at least 60% identity to the nucleic acid having the sequence as shown in SEQ ID NO:1 or is capable of hybridizing to the polynucleotide having the sequence as shown in SEQ ID NO:1 under conditions of intermediate to high stringency, or is complementary to the polynucleotide having the sequence as shown in SEQ ID NO:1. In another embodiment, the polynucleotide has the sequence as shown in SEQ ID NO:1.

The present invention also relates to methods for screening for mutants exhibiting a hyper branching phenotype and which are capable of secreting heterologous protein.

Accordingly, the present invention provides a method for the identification of hyperbranching mutants which comprise the steps of obtaining fungal mutants, subjecting said mutants to selection under desired conditions, and identifying mutants having the desired phenotypes. In one embodiment, the identification comprises selecting for hyphal growth. In yet another embodiment, identification comprises selecting for mutants capable of secreting protein. In another embodiment, the selection comprises growth and/or secretion of heterologous proteins at a restricted temperature.

#### **Brief Description of the Drawings**

Figures 1A-1D illustrates the nucleic acid (SEQ ID NO:1, hbrA) and amino acid (SEQ ID NO:2) sequence for HbrA.

Figures 2A-2B illustrates an amino acid alignment of the amino acid sequence for hbrA; A. fumigatus (afvac); rat (ratvac); yeast slp gene (slp1\_yeast); C.elegans (slp1\_ceel). Figure 3 illustrates amylase secretion by hbr/creA mutants.

### Detailed Description of the Invention

#### **Definitions**

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As used herein, the phrase "protein associated with hyphal growth" refers to a protein which is capable of modulating hyphal growth in fungus. Illustrative of such proteins are the proteins HbrA 1-9 disclosed herein in the Examples. The term "HbrA" refers to the amino acid sequence as shown in SEQ ID NO:2. The present invention encompasses proteins associated with hyphal growth in fungus having at least 70%, at least 75%, at least 80%, at least 85%, at least 90% or at least 95% identity to the amino acid sequence as disclosed in SEQ ID NO:2. Percent identity at the nucleic acid level is determined using the FastA program and percent identity at the amino acid level is determined using the TFastA both of which use the method of Pearson and Lipman (PNAS USA, 1988, 85:2444-2448). The present invention also encompasses mutants, variants and derivatives of HbrA as long as the mutant, variant or derivative is capable of modulating hyphal growth in fungus.

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As used herein, "nucleic acid" refers to a nucleotide or polynucleotide sequence, and fragments or portions thereof, and to DNA or RNA of genomic or synthetic origin which may be double-stranded or single-stranded, whether representing the sense or antisense strand. As used herein "amino acid" refers to peptide or protein sequences or portions thereof.

The terms "isolated" or "purified" as used herein refer to a nucleic acid or amino acid that is removed from at least one component with which it is naturally associated.

As used herein, the term "heterologous" when refering to a protein associated with hyphal growth refers to a protein that does not naturally occur in a fungal cell. The term "homologous" when refering to a protein associated with hyphal growth refers to a protein native or naturally occurring in the fungus. The invention includes fungal host cells producing the homologous protein associated with hyphal growth at higher copy number than found in the naturally occurring fungal host and produced at a higher copy level via recombinant DNA technology.

As used herein, the term "overexpressing" when referring to the production of a protein in a host cell means that the protein is produced in greater amounts than its production in its naturally occurring environment.

#### Description of the Preferred Embodiments

The present invention relates to the identification of HbrA in *A. nidulans*. The mutation of HbrA, referred to herein as HbrA2, was assigned to chromosome VII by parasexual analysis (*Aspergillus*: 50 Years On (1994) vol 20, ed S.D. Martinelli & J.R. Kinghorn pp. 41-43). At 37°C, mutant *hbrA*2, unlike wild-type *A. nidulans*, fails to secrete recombinantly expressed *M. meihei* protease. The translated sequence of the *hbrA*2 gene shows significant identity with the yeast SLP/VPS33 Sec1 gene product. Available evidence indicates that SLP/VPS33 Sec1 encodes a protein essential for vacuolar protein sorting. SLP1 mutants fail to direct proteins to the vacuoles, and they are sent along a default pathway to the cytoplasmic membrane. The exact nature and function of the SLP1/VPS33 Sec1 protein is unknown, but it is a member of the SEC1 family, and may be a membrane associated protein involved in directing vesicles to vacuoles. Deletion of VPS33 in yeast in not lethal, but leads to slow growth, temperature sensitivity, and loss of vacuoles as revealed by staining light and electron microscopy. Fluorescence microscopy has shown that like SLP1/VSP33 mutants in yeast, HbrA2 is defective in vacuole assembly at the non-permissive temperature.

The mutation HbrA2 does not appear to affect growth of *A. nidulans* at 26°C, but results in a hyperbranching, restricted growth phenotype at 42°C. The hyperbranching

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phenotype shows extensive branching in the apical compartment, unlike the wild-type *A.nidulans*. The mutant grows slowly at the non-permissive temperature giving rise to highly compact colonies on agar media. *Mucor meihei* protease was transformed into wild-type *A.nidulans* and crossed into the *hbrA2* mutant. The *hbrA2* mutant comprising the heterologous nucleic acid encoding the *M.meihei* protease was able to secrete the protease at 26°C. The *hbrA2* mutant was unable to secrete the protease at 37°C but was able to secrete the endogenous alpha amylase at temperatures greater than 37°C.

In view of the observation that *hbrA* mutants are incapable of producing foreign protein, it appears that genetically engineering fungal hosts to modulate the expression of proteins associated with hyphal growth, in particular, mutants HbrA1-9, would provide a means for enhancing the production of proteins or chemicals in the fungal host. In one aspect of the present invention, it would be desirable to increase expression of proteins associated with hyphal growth. In another aspect of the present invention, it would be desirable to decrease or eliminate expression of proteins associated with hyphal growth by means known to the skilled artisan.

I. HbrA amino acid and hbrA nucleic acid sequences

The present invention provides the amino acid (SEQ ID NO:2) HbrA and nucleic acid (SEQ ID NO:1) sequence for hbrA. The present invention encompasses amino acid variants having at least 70% identity to the amino acid having the sequence as shown in SEQ ID NO:2 as long as the variant is capable of modulating hyphal growth. Percent identity at the nucleic acid level is determined using the FastA program and percent identity at the amino acid level is determined using the TFastA both of which use the method of Pearson and Lipman (PNAS USA, 1988, 85:2444-2448). Alternatively, identity is determined by MegAlign Program from DNAstar (DNASTAR, Inc. Maidson, WI 53715) by Jotun Hein Method (1990, Method in Enzymology, 183: 626-645) with a gap penalty = 11, a gap length penalty = 3 and Pairwise Alignment Parameters Ktuple = 2. As the skilled artisan will readily recognize, a variety of polynucleotides can encode HbrA. The present invention encompasses all such polynucleotides. HbrA, and other polynucleotides encoding proteins associated with hyphal growth, may be obtained by standard procedures known in the art from, for example, cloned DNA (e.g., a DNA "library"), genomic DNA libraries, by chemical synthesis once identified, by cDNA cloning, or by the cloning of genomic DNA, or fragments thereof, purified from a desired cell. (See, for example, Sambrook et al., 1989, Molecular Cloning, A Laboratory Manual, 2d Ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York; Glover, D.M. (ed.), 1985, DNA Cloning: A Practical Approach, MRL Press, Ltd., Oxford, U.K. Vol. I, II.) Nucleic acid

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sequences derived from genomic DNA may contain regulatory regions in addition to coding regions. Whatever the source, the isolated polynucleotide encoding the protein associated with hyphal growth can be molecularly cloned into a suitable vector for propagation of the gene.

In the molecular cloning of the gene from genomic DNA, DNA fragments are generated, some of which will encode the desired gene. The DNA may be cleaved at specific sites using various restriction enzymes. Alternatively, one may use DNAse in the presence of manganese to fragment the DNA, or the DNA can be physically sheared, as for example, by sonication. The linear DNA fragments can then be separated according to size by standard techniques, including but not limited to, agarose and polyacrylamide gel electrophoresis and column chromatography.

Once the DNA fragments are generated, identification of the specific DNA fragment containing the gene may be accomplished in a number of ways. For example, a polynucleotide encoding a protein associated with hyphal growth or its specific RNA, or a fragment thereof, such as a probe or primer, may be isolated and labeled and then used in hybridization assays to detect related genes. (Benton, W. and Davis, R., 1977, Science 196:180; Grunstein, M. And Hogness, D., 1975, Proc. Natl. Acad. Sci. USA 72:3961). Those DNA fragments sharing substantial sequence similarity to the probe will hybridize under stringent conditions.

Also included within the scope of the present invention are fungal microorganism polynucleotide sequences that are capable of hybridizing to the nucleotide sequence of SEQ ID NO:1 under conditions of intermediate to maximal stringency. Hybridization conditions are based on the melting temperature (Tm) of the nucleic acid binding complex, as taught in Berger and Kimmel (1987, Guide to Molecular Cloning Techniques, Methods in Enzymology, Vol 152, Academic Press, San Diego CA) incorporated herein by reference, and confer a defined "stringency" as explained below.

"Maximum stringency" typically occurs at about Tm-5°C (5°C below the Tm of the probe); "high stringency" at about 5°C to 10°C below Tm; "intermediate stringency" at about 10°C to 20°C below Tm; and "low stringency" at about 20°C to 25°C below Tm. As will be understood by those of skill in the art, a maximum stringency hybridization can be used to identify or detect identical polynucleotide sequences while an intermediate or low stringency hybridization can be used to identify or detect polynucleotide sequence homologs.

The term "hybridization" as used herein shall include "the process by which a strand of nucleic acid joins with a complementary strand through base pairing" (Coombs J (1994) Dictionary of Biotechnology, Stockton Press, New York NY).

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The process of amplification as carried out in polymerase chain reaction (PCR) technologies is described in Dieffenbach CW and GS Dveksler (1995, PCR Primer, a Laboratory Manual, Cold Spring Harbor Press, Plainview NY). A nucleic acid sequence of at least about 10 nucleotides and as many as about 60 nucleotides from SEQ ID NO:1 preferably about 12 to 30 nucleotides, and more preferably about 20-25 nucleotides can be used as a probe or PCR primer.

#### **Expression Systems**

The present invention provides host cells, expression methods and systems for the production of desired proteins in host fungus. Once nucleic acid encoding a protein associated with hyphal growth is obtained, recombinant host cells containing the nucleic acid may be constructed using techniques well known in the art. Molecular biology techniques are disclosed in Sambrook et al., Molecular Biology Cloning: A Laboratory Manual, Second Edition (1989) Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY (1989). Nucleic acid encoding proteins associated with hyphal growth and having at least 60% identity to *hbrA* is obtained and transformed into a host cell using appropriate vectors. A variety of vectors and transformation and expression cassettes suitable for the cloning, transformation and expression in fungus are known by those of skill in the art.

Typically, the vector or cassette contains sequences directing transcription and translation of the nucleic acid, a selectable marker, and sequences allowing autonomous replication or chromosomal integration. Suitable vectors comprise a region 5' of the gene which harbors transcriptional initiation controls and a region 3' of the DNA fragment which controls transcriptional termination. These control regions may be derived from genes homologous or heterologous to the host as long as the control region selected is able to function in the host cell.

Initiation control regions or promoters, which are useful to drive expression of the protein associated with hyphal growth in a host cell are known to those skilled in the art. Virtually any promoter capable of driving these proteins is suitable for the present invention. Nucleic acid encoding the protein is linked operably through initiation codons to selected expression control regions for effective expression of the protein. Once suitable cassettes are constructed they are used to transform the host cell.

General transformation procedures are taught in Current Protocols In Molecular Biology (vol. 1, edited by Ausubel et al., John Wiley & Sons, Inc. 1987, Chapter 9) and include calcium phosphate methods, transformation using PEG and electroporation. For Aspergillus and Trichoderma, PEG and Calcium mediated protoplast transformation can be used (Finkelstein, DB 1992 Transformation. In Biotechnology of Filamentous Fungi.

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Technology and Products (eds by Finkelstein & Bill) 113-156. Electroporation of protoplast is disclosed in Finkelestein, DB 1992 Transformation. In Biotechnology of Filamentous Fungi. Technology and Products (eds by Finkelstein & Bill) 113-156. Microprojection bombardment on conidia is described in Fungaro et al. (1995) Transformation of Aspergillus nidulans by microprojection bombardment on intact conidia. FEMS Microbiology Letters 125 293-298. Agrobacterium mediated transformation is disclosed in Groot et al. (1998) Agrobacterium tumefaciens-mediated transformation of filamentous fungi. Nature Biotechnology 16 839-842.

Host cells which comprise the sequence for *hbrA* and express the protein may be identified by a variety of procedures known to those of skill in the art. These procedures include, but are not limited to, DNA-DNA or DNA-RNA hybridization and protein bioassay or immunoassay techniques which include membrane-based, solution-based, or chipbased technologies for the detection and/or quantification of the nucleic acid or protein. For production of a desired protein in a fungal host cell, an expression vector comprising at least one copy of nucleic acid encoding a desired protein is transformed into the recombinant host cell comprising nucleic acid encoding a protein associated with hyphal growth and cultured under conditions suitable for expression of the protein. Examples of desired proteins include enzymes such as hydrolases including proteases, cellulases, amylases, carbohydrases, and lipases; isomerases such as racemases, epimerases, tautomerases, or mutases; transferases, kinases and phophatases along with proteins of therapeutic value. Alternatively, it may be advantageous to down-regulate or mutate proteins associated with hyphal growth in order to reduce the viscosity in the fermentor.

III Vector Sequences

Expression vectors used in expressing the *hprA* in fungal cells or the desired protein in fungal cells comprise at least one promoter associated with the protein which promoter is functional in the host cell. In one embodiment of the present invention, the promoter is the wild-type promoter for the protein and in another embodiment of the present invention, the promoter is heterologous to the protein, but is still functional in the fungal host cell. In one preferred embodiment of the present invention, nucleic acid encoding the protein is stably integrated into the microorganism genome.

In a preferred embodiment, the expression vector contains a multiple cloning site cassette which preferably comprises at least one restriction endonuclease site unique to the vector, to facilitate ease of nucleic acid manipulation. In a preferred embodiment, the vector also comprises one or more selectable markers. As used herein, the term

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selectable marker refers to a gene capable of expression in the host which allows for ease of selection of those hosts containing the vector.

#### IV. Assay of the activity of proteins associated with fungal growth

The results shown in Examples I and II illustrate the use of a temperature based screen to identify mutants which effect fungal branching. The unexpected advantage of using such a temperature based screen is the ability to identify HbrA mutants or mutants of proteins associated with hyphal growth having a differential effect on the export of native or endogenous genes vs the export of recombinantly introduced heterologous protein. This type of screening method facilitates the isolation of strains which are capable of increased secretion of heterologous protein. Therefore, the present invention also provides a method for the identification of hyper-branching mutants which enhance protein secretion comprising the steps of obtaining fungal mutants, subjecting said mutants to selection under desired conditions, and identifying the desired mutants. In one embodiment, the identification comprises selecting for hyphal growth. In another embodiment, the selection comprises growth and/or secretion of heterologous proteins at a restricted temperature.

#### Examples

#### Example I

This example illustrates the isolation of the hbrA gene. In order to isolate the hbrA gene, DNA was prepared from pooled cosmids of the chromosome-sorted cosmid library of wild-type DNA from A. nidulans obtained from FGSC (Funal Genetic Stock Center, Department of Microbiology University of Kansas Medical Center, Kansas City, KS 66160). 5 pools of 20 cosmids each were used in transformation experiments. In order to assess transformation efficiency, an hbrA2, argB double mutant was used as a recipient for cotransformation using a mixture of cosmid DNA and transforming vector Arp, which carries the argB gene and a replicating sequence. After transformation, protoplasts were regenerated and selected on medium lacking arginine at 42°C. One of the cosmid pools gave rise to a few strongly growing, normally conidiating colonies in a background of Arg+ Hbr- transformants. The pool was subdivided into 4 pools of 5 cosmids, and transformation repeated. A single cosmid was isolated which was able to complement the hbrA2 mutation, restoring wild-type growth. Sub-cloning of the cosmid led to identification of an EcoRI fragment carrying the transforming sequence. The EcoRI/BamHI fragments failed to complement the mutation suggesting that the BamHI site lies within the hbrA gene. The fragment was isolated and subjected to nucleic acid sequencing. The nucleic acid and amino acid sequence for the hbrA gene is shown in Figures 1A-1D. Table I shows

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protease activity for Hbr2, as well as other identified hyper-branching mutants at the permissive and non-permissive temperatures.

Table I

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v	Strain	Mean Protease Activity (units/gram of blomass) at 26C 48 hrs 72 hrs			tease Activity of biomass) at (37C) 72 hrs
10	Wild-type	963+/- 57	703+/-12	380+/-44	339+/- 40
	HbrA2	857+/-18	1237+/-155	0+/- 0	0+/-0
	Hbr3	689+/-76	1194+/-234	0+/-0	0+/-0
15	Hbr6 Hbr8	0+/-0 0+/-0	1892+/-122 2165+/-156	0+/-0 0+/-0	0+/-0 487+/-10

These findings indicate that a previously uncharacterized filamentous fungal gene hbrA plays a role in heterologous protein export.

#### Example 2

This Example describes the characterization of hyperbranching mutants of A. nidulans. Below is Table II which shows the chromosomal location of the hbr mutants.

	hbr Mutant	Chromosomal location
	hbr1	1
	hbrA2	VII
30	hbr3	i
	hbr4	tii
	hbr5	VIII
	hbr6	Ш
	hbr7	111
35	hbr8	1
	hbr9	III

All mutations were recessive and unlinked to each other and represent previously uncharacterized mutations which effect fungal hyperbranching and protein secretion. The ability of hbrA2 mutant to secrete the endogenous protein alpha amylase at 37°C was examined by growing the hbrA2:creA- double mutant on petri dishes with starch as the sole carbon source (the CreA gene is a negatively acting regulator of carbon catabolism repression. Mutations of CreA (CreA-) causes carbon catabolism derepression of enzymes such as alpha amylase). The hbrA2:creA- double mutant like the hbrA+:creA- was shown to be capable of secreting the endogenous protein alpha amylase, see Figure 3.

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These results indicate the hbrA gene unexpectantly plays a role in heterologous protein secretion.

The hbr3 mutant, like the hbrA2 mutant, produces slightly higher M. meihei protease than the wild-type at 26°C. At 37°C, the hbr3 mutant like the hbrA2 mutant does not produce the M. meihei protease. The hbrA2 mutation is located on chromosome VII, the hbr3 mutation is located on chromosome I. These results Indicate that the hbr3 gene product also plays a role in heterologous protein export. Therefore, modulation of the expression of the wild-type hbr3 gene product would appear to be advantageous in increasing heterologous protein export.

The hbr6 and hbr8 mutations which are located on chromosomes III and I respectively, produce significantly higher levels of M. meihei protease than the wild-type at 26°C and would appear to increase the secretion of heterologous protein in a filamentous fungus grown in the temperature range around 26°C. Therefore, modulation of expression of the wildtype hbr6 and hbr8 gene products would also appear to have utility in increasing heterolgous protein export. Mutant versions of the hbr6 and hbr8 genes have no or significantly less M. meihei secretion than the wild-type as shown by Table III.

#### Table III

20	Strain	Mean Protease Activity (units/gram of biomass) at 26C 48 hrs 72 hrs			tease Activity of biomass) at 37C 72 hrs
	Wild-type	963+/- 57	703+/-12	380+/-44	339+/- 40
25	hbr5 hbr7	46+/-60 0+/-0	1152+/-133 1098+/-53	533+/-53 580+/-60	1648+/-797 1581+/-660
30	hbr4	844+/-114	1688+/-67	343+/-26	260+/-15
	hbr9	0+/-0	268+/-16	0+/-0	1562+/641

Table II illustrates that M. meihei protease secretion in the hbr5 and hbr7 mutants yields slightly more protease at 26°C after 72 hours compared to the wild-type, and significantly more protease at 72 hours at 37°C.

The hbr4 mutant produced significantly more M. meihei protease than the wild-type after 72hours at 26°C but significantly less protease after 72 hours at 37°C. However, the hbr4:creA- double mutant produced significantly higher levels of alpha amylase/unit area fungal colony that the wild-type strain containing only the creA- mutation. These results indicate a significant role for the hbr4 gene product not only in terms of fungal morphology

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The hbr9 mutation exhibited poor expression of M. meihei protease at 26°C, but

increasing native protein secretion but also a role for this gene product in heterologous protein export.

significantly higher levels of M. meihel protease and alpha amylase/ unit area fungal

5 colony than the wild-type.

### Claims

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We claim:

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1. An isolated protein associated with hyphal growth in fungi having at least 70% identity to the amino acid sequence as disclosed in SEQ ID NO:2.

- 2. The protein of Claim 1 having the amino acid sequence as disclosed in SEQ ID NO:2.
- An isolated polynucleotide encoding the amino acid having the sequence as shown in SEQ ID NO:2.
- 4. The isolated polynucleotide of Claim 3 having at least 60% identity to the polynucleotide have the sequence as shown in SEQ ID NO: 1, or is capable of hybridizing to the polynucleotide having the sequence as shown in SEQ ID NO:1 under conditions of intermediate to high stringency, or is complementary to the polynucleotide having the sequence as shown in SEQ ID NO:1.
- 5. The Isolated polynucleotide of Claim 4 having the nucleic acid sequence as disclosed in SEQ ID NO:1.
  - 6. An expression vector comprising the polynucleotide of Claim 3.
  - 7. A host cell comprising the expression vector of Claim 6.
  - 8. The host cell of Claim 7 that is a filamentous fungus.
- 9. The host cell of Claim 8 wherein said filamentous fungus includes Aspergillus, Trichoderma, Mucor and Fusarium.
- 10. A method for producing a desired protein in a fungus comprising the step of, culturing a recombinant fungus comprising a polynucleotide encoding the desired protein under conditions suitable for the production of said desired protein, said recombinant fungus further comprising a polynucleotide encoding a protein associated with hyphal growth in said fungus said protein having at least 70% identity to the amino acid sequence as disclosed in SEQ ID NO:2.

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5	11. The method of Claim 10 further comprising the step of recovering said desired protein.
10	12. The method of Claim 10 wherein said polynucleotide encoding a protein associated with hyphal growth is homologous to said fungus said polypeptide being present in copy number greater than found in the naturally occurring fungus.
15	13. The method of Claim 10 wherein the polynucleotide encoding a protein associated with hyphal growth is heterologus to said fungus and has been recombinantly introduced into said fungus.
20	14. The method of Claim 10 wherein said polynucleotide encoding a protein associated with hyphal growth in said fungus comprises a replicating plasmid.
25	15. The method of Claim 10 wherein said polynucleotide encoding a protein associated with hyphal growth in said fungus is integrated into the fungal genome.
30	16. The method of Claim 10 wherein said protein associated with hyphal growth has the amino acid sequence as shown in SEQ ID NO:2.
	17. The method of Claim 10 wherein said polynucleotide encoding a protein associated with hyphal growth has 60% identity to the polynucleotide having the sequence as shown in SEQ ID NO:1, or is capable of hybridizing to the polynucleotide having the
35	sequence as shown in SEQ ID NO:1 under conditions of intermediate to high stringency, or is complementary to the polynucleotide having the sequence as shown in SEQ ID NO:1.
40	18. The method of Claim 15 wherein said polynucletoide has the nucleic acid sequence as shown in SEQ ID NO: 1.
	19. The method of Claim 10 wherein said fungus is a filamentous fungus.
45	20. The method of Claim 19 wherein said filamentous fungus includes  Aspergillus, Trichoderma, Mucor and Fusarium species.

nidulans, A. oryzai and A. fumigatus.

The method of Claim 20 wherein the Aspergillus includes A. niger, A.

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5 A method for producing a recombinant fungus comprising a polynucleotide 22. encoding a protein associated with hyphal growth comprising the steps of: (a) obtaining a polynucleotide encoding said protein associated with; (b) introducing said polynucleotide into said host cell; and 10 (c) growing said host cell under conditions suitable for the production of said protein associated with hyphal growth. 23. The method of Claim 22 wherein said host cell includes filamentous fungus. 15 The method of Claim 23 wherein said filamentous fungus includes 24. Aspergillus, Trichoderma, Mucor, and Fusarium. 20 The method of Claim 24 wherein said Aspergillus species includes A. niger, A. nidulans, A. oryzae and A. fumigatus, 26. The method of Claim 22 wherein said polynucleotide has at least 60% 25 identity to the nucleic acid having the sequence as shown in SEQ ID NO:1. 27. A method of identifying hyper-branching fungal mutants comprising the 30 steps of obtaining fungal mutants, subjecting said mutants to selection under desired conditions, and identifying the desired mutants. 28. The method of Claim 27 wherein said identifying comprises selecting for 35 hyphal growth. 29. The method of Claim 27 wherein the selection comprises growth and secretion of heterologous proteins at a restricted temperature. 40 30. A method for producing a desired protein in a fungus comprising the step of culturing a recombinant fungus comprising a polynucleotide encoding the desired protein under conditions suitable for the production of said desired protein, said recombinant 45 fungus comprising a mutation in an endogenous nucleic acid encoding a protein associated with hyphal growth said mutation resulting in the inhibition of the production by said fungus

of the protein associated with hyphal growth.

The method of Claim 30 wherein said protein associated with hyphal growth

has at least 70% identity to the amino acid sequence as disclosed in SEQ ID NO:2.

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10	30	50
GATCACCAGGAATTG	CGTTGCCTGATGCATGGTTGGGAG	GGCCGCCGAGGTCCACGCCAG
70	90	110
GTGGTGGGGGTGCTA	TACCGTGCTGCGCTTTTGCCCTCG	TGTAAGGGTCAGCAGGAATCG
130	150	170
GTTTCGCGTAAGGAT	TCGCTTCGGCAGGAGGGCTCTTGT	TCTTCGACCTCGATCCAAAGA
190	210	230
GGGCGCGGGTTGG	AGGAATCGTCGTCGCCGGCGTCTG	ACGACTTTTTGAGGCCGAATC
250	270	290
GCTTCATAGCGTATT	TTAGCTAGAATACTTCGCCGAAAc	CAGCGTAGGAATALTAGAGTG
310	330	350
AAAATAATAAATTGA	GAGGCTATTTATGATTGACTGAGA	ATTGAAGAGAGGGAAGGGAA
370	390	410
GGAGGGAGGGAGCG	AAGATGTTAAGTGTCAGGGGAGCA	GCAGCGGCAAAAGTGTCAAGA
430	450	470
CGCTCCTGAGACTCA	AAGGCAGCTATGTAATCATGATAC	ACATAGTTGTGCTGCAATTCT
490	510	530
GGCTATCAGTGAGTA	TTTTACCGTATGATTACTCACCAA	TTCGACTCCACTAAGCCGAAA
550	570	590
GAAGCTAGCGGGGAT	GGCTGGACCCTTCTAAGCCTCAAC	TGAGGGCGGTGCCGCAGTCAA
610	630	650
ACGTCAACTGCTCCC	ACCCCATGCTTCGTATAAGGTAGC	CATGGCACCATTCCCTGGGTC
670 •	690	710
TGATGCCGACAATAT	CAAGGACAAGGCCCGTAAAGGCTT	GCTGAATCTTCTCGAAGGCGT

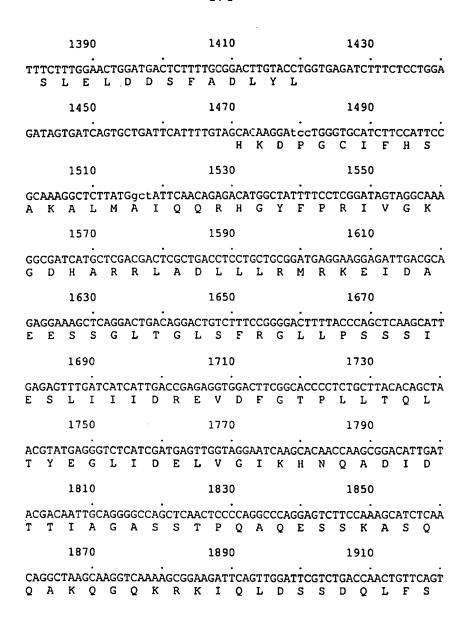
# FIG.\_1A

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730 750 770 GAGTAAGGCTCCTAGTTGGCACTGTTTCTGGTTCTAGCCTGATTCATTACCTCGATCTAG 790 810 830 GTCCGTGGGAAGAAGAACCTGGTGATTAGCCAGGGGCTTGCTGGGCCCGTCGGGCTTTTT 870 850 890 930 AATGTCGACTCTTCTCAGCGCAATGTGGTATTTCTAGCGTACGCCGAAAAGATCCGCCAG GTGCGGGCAGTGGCAGGTATGTCATGATCTTTATCCACCTTTGATTTACATACCCAAATG 1030 1050 ACTGTAAATGCGAAGGCTCCTTGCTATCGCGCTTGCTGGGAGCATTAAAGTTACGCAGAC 1090 1110 1130 TTCTTCTCCACTCTGCGTAATCAGTCAAGCTCCCTATATTGAAACTTCGTTTAGCAGCTT 1150 1170 1190 ATCCCTAAGGCTTTCTTTCTCTGCCTCGTATGACTGAATGCCATCAGAATAAGCTGACAA GTTTTACAGAGCAGATCCAAAGGCTTCAACGCAACAGCAGTATAGACCATGAATTTTCCA HEFSI 1270 1290 1310 TCTTTTGGGTTCCAAGACGGACCCTCGTAAGCAATAACATCCTAGAGAGCGCAGGCATCA FWVPRRTLVSNNILESAGII 1330 1350 TTGGAGATGTGAGCATCGCTGAGCTGCCTCTTTACTTTTTCCTCTAGAGCAGGACGTTC G D V S I A E L P L Y F F P L E Q D V L

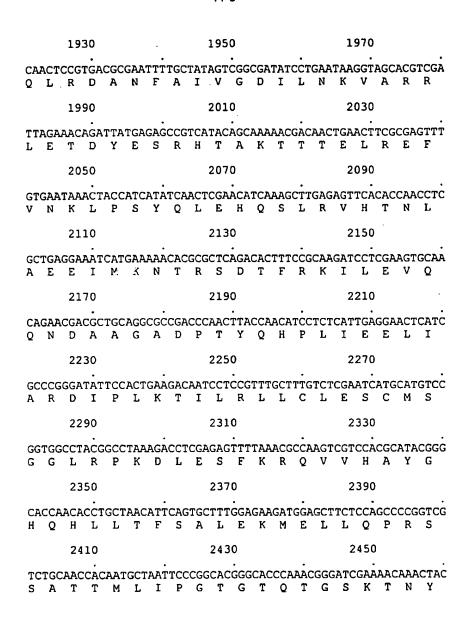
# FIG.\_1B

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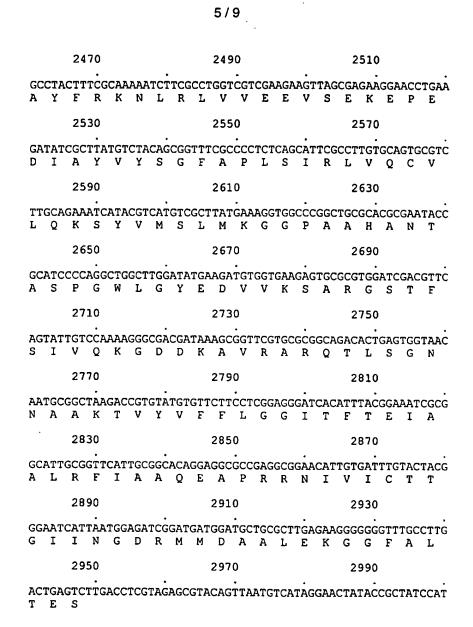


### FIG.\_1C

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### FIG.\_1D



# FIG.\_1E

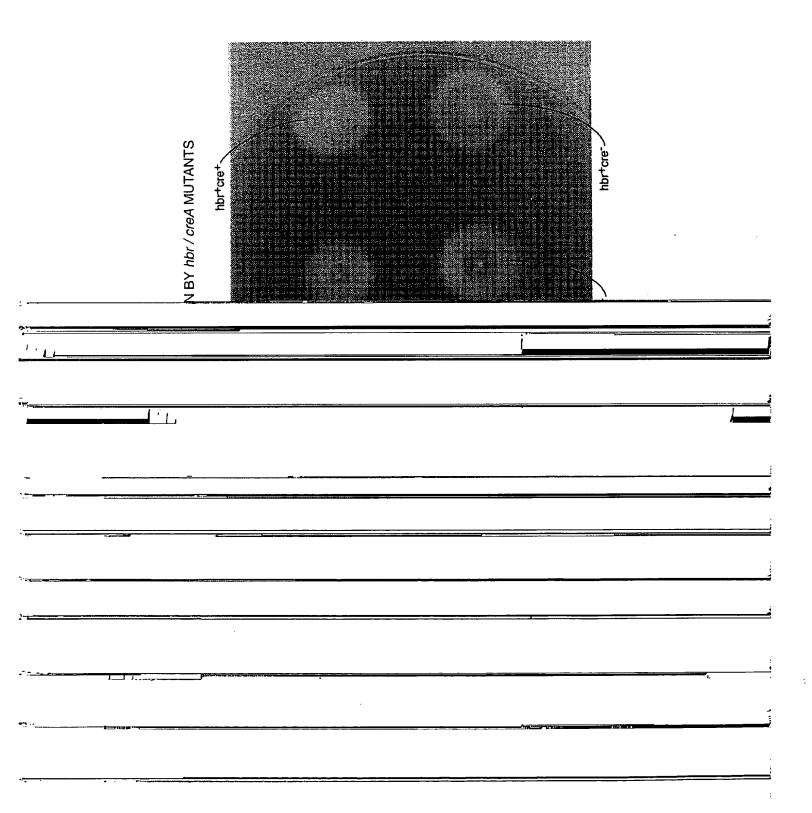
		~~ <del>~</del>	m /o =	
32000	8 8 114 116 93	42 42 149 176 134	91 91 198 236 191	151 151 247 296 296
OYSLLKEHEV TFQDLTQSTP SANEYSRNLL	··HEFSIFWV ··HEFSIFWL PTRDPHILFV NLSKIPHVWFL	YFFFLEQ YFVPLEQ DLIPFDG DLPFPQIDE CWLPRDG	FPRIVGKGDH FPRIIGKGDH IPQIFGHGEC ITNQVSIGNE PMSIYÄRGKW	TPLLTQLTYE TPLLTQLTYE TPLASQLTYE TPLTQLTYA
YLTGPFGLIA SVLPVLNSLL LNWEGTSEIK	ENVLSEDRRG ENVLSEDRRG PQLLDAAQKH ANHSIETKKS	AEPL EEYSL LSNEYTYPN RLESVKEIPL	H TBA THORY IN THE THE THORY IN THE THE THORY IN THE	iiidrevdfg iiidrmvdfg iillernvdli iillentdpi
AGSKAIVWDE QNDEVLVVQP NEDRDDAAAI	DOURTSLRIP ASVLAAHGVV	AGIIGD.VSI AGIIGD.VNI LGVLGSFIYR PNVIESQLKE DNVRNDSYWE	MAIQOR MDIQOR MTLOAL (TLOEDMUNSL NOLIDMCRGR	LLPSSSIESL LLPSASTESL DNL RTNCGLEMDL IEGLLKINR
RELREFLDKC GLCATLNEIS	FLVRPRLELM SVPQMDLIFL VMHRVNLFAG	FDLNSKKTHF	GCIFHSAKAL GCIFLAAKAL TSLYHAAKĞI LRSLQSATES THLHKCAVAL	SGLTGLSFRG SGLRGPSARG SVFPVF LFIKGTLYGE
NLNVLREAVR TKKFSLTNAD	 LPAADVKNII LSDDLPSALG KTIVWDRDRS	LES LED LKD EDTEQWQKDG	FADLYLHKDP FGDLYLHKDP FKECYLEGDQ MENÜMÖYYPN AARLIÄNGDW	mrkeidaees mrkeldaees mkreftgison ridehqtend itrnsaea@s.
MAAHLSYGRV	EKMFTLKGSR VRKITLEDDO FSVLDSLDGN	PRRTEVSNNI PRRTEVSNKI PRSLÄGEOR RWKPSFONTL Ä: PTMVSLDL	DVLSLELDDS DVLSLELDDS DŽILSMESESA NVLLTHSLYN ECLSLSSPQI	arrladilir arrladilir arqvanMMvr skrcshilkk asdvakMMGkk
hbra afvac ratvac slp1_yeast slp1_caeel	hbra afvac ratvac slp1_yeast slp1_caeel	hbra afvac ratvac slp1_yeast slp1_caeel	hbra afvac ratvac slp1_yeast slp1_caeel	hbra afvac ratvac slp1_yeast slp1_caeel

FIG.\_2A

D. BSDQLFSQ 209 D. SSDQLFSQ 210 N. SSDQLFSQ 210 NYKEDKÄWND 328	QSLRVHINLA 269 QSLRVHINLA 270 GSLANHISKA 359 AFLKNHTILS 388 KMČEVHMRLA 352	IPLKTĪLRLL 321 VPLKTVLRLU 322 HPLIKVLRLV 411 VDQKKĪLRLI 445 SPLNAVLRLI 403	TMLIPGIGIQ 381 TMLIPTAGAQ 382 DEHITTISKS 504 KMQCEYA 460	RLVQCVLQKS 432 RLVQCVL 430 RLAQLL 504 RLVQMLYDRS 564 WMLE 502
KOGOKŘKIQU KOGOKŘKIQU TŘAKKLQU IKIRKKKOK DPFQEKEVYL	NKLPSYQLEH (NKLPSYQLEH (SQLPHQAAR CDSLGSLQQRQ)	LIARD J LIARD J CIAQR H KILELQYQYE LIFDA	Rillo Prssat Selo Prssaa Glik. Gerrakti. Gerrakger Getrekgege	VYSGFAPLSI I VYSĞFAPLSI I VYSGÜAPLSV I AYSGVVPLTM I
ESSKASQQA. ESSKAPQQTL GGGGGKDLP. EMEFKNEKDG	KTTTELREFV KTASELREFV KTVGEIKÖFV ESVHEIKEFV MSVAEYSVÜÖV	PTYQHPLIEE PTYQHDTIEE TDKVNNYIED .NTLNNDIED SDKAIPLIED	LLTFSALEKM ILTLNNLEKM ILTLNNLEKA CEQLESLA ELI LNKVLK图贷KM	SEKEPEDIAY SEEDPNDIAY NEQNPTDISY EMODFSEATE
AGASSTPQAQ VGANSVPQAQ PPEKFAPKQ	ETDYESRHTA ESEYETRHAA SAADEERHNA QTQYDTRKA RDDEQEGGE	VÖGNDAAGAD VÖGNNAAGTD VEGEFNSGID HELELLMG	QVVHAYGHQH QIVHAYGHQH EILQTYGYEH FMIDSMGIEK IVNQSYGSSA	V
HNQADIDTTI NNQADVDTAI NSYVK SG	DILNKVARRE DILNKVARRE SVLSKKAKIË PQLNKËAKEL IBASKVLAEË	TFRKILE IFRKULE DFFDKLT EYGSFNKILE VYCKQSDSÄK	RPKDLESFKR RSPDLENFK KOKUTYKR REKDYEYLR KPSVLQHYRR	RKNLRLVVEE RKNLRLLVEE RKTLRLWMOD SQWENTVPIE KKNHOMLPEE
GLIDELVGIK GLIDEFVGIK GLIDEIYGIQ GLÜDELYEÜN GLÜDEIYGI	LRDANFAIVG KRDANFAIVG KRDKNFNAVG LKELNFGSKIG LKHSKINAVS	EEIMKNTRSD ÖEIMRNTRSD ELIKDVTTSE SDÜLKVVETE EMIQSH	CLESCMSGGL CLESCMSGGL CLESCKSGL CLESCKNSGL SVHSLTCGGL	TGSKTNYAYF PGTKTNYNYL TGGRNNYPTI TRLQKËYRYI
hbra afvac ratvac slp1_yeast slp1_caeel	hbra afvac ratvac slp1_yeast slp1_caeel	hbra afvac ratvac slp1_yeast slp1_caeel	hbra afvac ratvac slp1_yeast slp1_caeel	hbra afvac ratvac slp1_yeast slp1_caeel

492 430 544 624 530	534 430 586 684 567		
hbra yvmslmkggp aahantaspg wlgyedvvks argstfsivo kgddkavrar Qtlsgnnaajä 492 afvacsrpg wrsieevlri lpgphfeero plptgvokkr QPgenr 544 spt_yeast ilfhnyssoo pfilsreprv sqtedlieol ygdshaiees iwvpgtitkk inasiksnnr 624 slp1_caeel	ICTTGÄINGD ISTTKLINGS IIADGLINGT ITTSALITGD		
KGDDKAVRAR PLPTGVQKKR IWVPGTITKK	AQEAPRRNIV OLEDGGTEYV GKKGINKRFI		
ARGSTFSIVQ  LPGPHFEERQ YGDSHAIEES	FLGGITFTEI AALRFIA AQEAPRRNIV FLGGVTFAEI AALRFLS QLEDGGTEYV FLGGVTMGEI AIMÄHLDKIL GKKGINKRFI VÄGGLTRSEÄ AIRENLPNVAL	·	70
WLGYEDVVKS WRSIEEVLRI SQTEDLIEQL	FLGGITFTEI FLGGVTFAEI FLGGVTMGEI VÄGGLTRSEÄ		בוני יינ
AAHANTASPG	ASIDGSNGTF HAAEDIALVV FLGGVTFAEI AALRFLS AQEAPRRNIV RSIDGSNGTF HAAEDIALVV FLGGVTMGEI AIMÄHLQKIL GKKGINKRFI GTCVF VÄGGLTRSEÄ AIR ENLPNVAL	GFALTES 551 7 430 7 4 691 7 691	
YVMSLMKGGP	RSIDGSNGTF	RMMDAALEKG GFALTES SWLEALWEKP F* RIMNSIS KLLNNITN	
hbra afvac ratvac slp1_yeast slp1_caeel	hbra atvac ratvac slp1_yeast i	hbra afvac ratvac slp1_yeast slp1_caeel	

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## INTERNATIONAL SEARCH REPORT

Internacional Application No PCT/US 00/07615

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	International Patent Classification (IPC) or to both national class	sification and IPC			
	SEARCHED				
IPC 7	ocumentation searched (classification system followed by classification C12N C12P C07K	ication symbols)			
Documentat	tion searched other than minimum documentation to the extent ti	nat such documents are included in the field	s searched		
Electronic da	ats base consulted during the international search (name of dat	a base and, where practical, search terms u	sed)		
BIOSIS	, EMBL, EPO-Internal, STRAND, CHE	M ABS Data, MEDLINE, WP1	I Data, PAJ		
C. DOCUME	ENTS CONSIDERED TO BE RELEVANT				
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X Furth	ner documents are listed in the continuation of box C.	X Patent family members are list	ted in annex.		
Special categories of cited documents:  'A' document defining the general state of the lart which is not considered to be of particular relevance.  'E' earlier document but published on or after the International filing date.  'L' document which may throw doubts on priority claim(e) or which is olded to establish the published on date of another citation or other special reason (as epselfled).  'O' document referring to an oral disclosure, use, exhibition or other means.  'P' document published prior to the international filing date but later than the priority date dalmed.  Date of the actual completion of the international search.		or priority date and not in conflict or close to understand the principle or invention.  "X" document of particular relevance; it cannot be concidered novel or car involve an inventive step when the "Y" document of particular relevance; it cannot be considered to involve at document is combined with one or menta, such combination being ob in the art.  "8," document member of the same path	"X" document of particular relevance; the claimed invention cannot be considered now or cannot be considered to involve an inventive step when the document is taken alone "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled		
	5 August 2000	28/08/2000			
Name and mailing address of the ISA  European Patent Office, P.B. 5818 Patentiaan 2  NL – 2280 HV Rijswijk  Tel. (-431-70) 340-2040, Tx. 31 651 epo ni,		Authorized officer			
	Fax: (+31-70) 340-3016	Lejeune, R			

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Accession AA784458, 8 February 1998 (1998-02-08) KUPFER D ET AL: "e4b02a1.r1 Aspergillus nidulans 24hr asexual developmental and vegetative cDNA lambda zap library Emericella nidulans cDNA clone e4b02a1 5', mRNA sequence" XP002144972 99.1% identity in 318 BP overlap with SEQ ID NO 1  P,X MEMMOTT S ET AL: "Abstract of Poster: 339. Morphological and genetic characterization of Hbr-2, a hyperbranching mutant of Aspergillus nidulans" 20TH FUNGAL GENETICS CONFERENCE, 'Online! 24 - 28 March 1999, XP002144968 Retrieved from the Internet:	4
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abstract W0 97 26330 A (NOVO NORDISK BIOTECH INC) 24 July 1997 (1997-07-24) abstract see examples see claims	27-29

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